

DM2 RESEARCH UPDATE

Giovanni Meola, MD, PhD

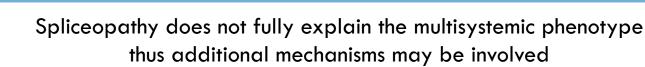


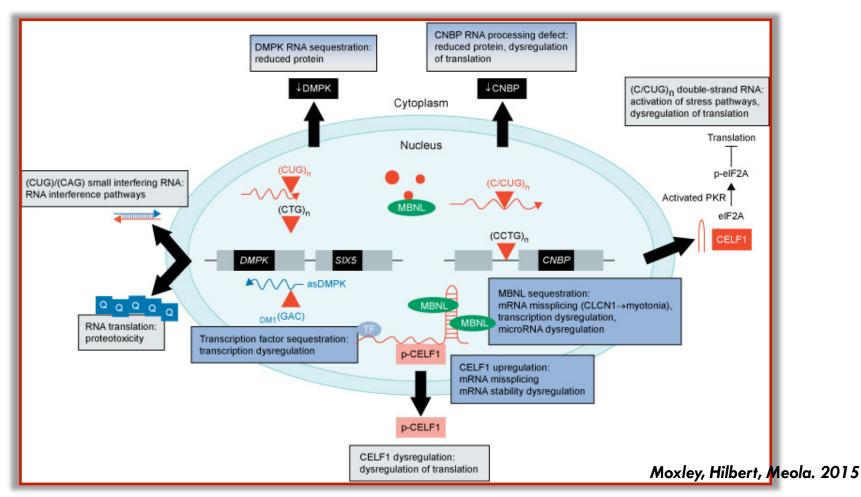




- PATHOGENESIS
- MODIFIER GENES
- MANAGEMENT
- MOLECULAR THERAPY
- TAKE HOME MESSAGE

Pathogenetic mechanism



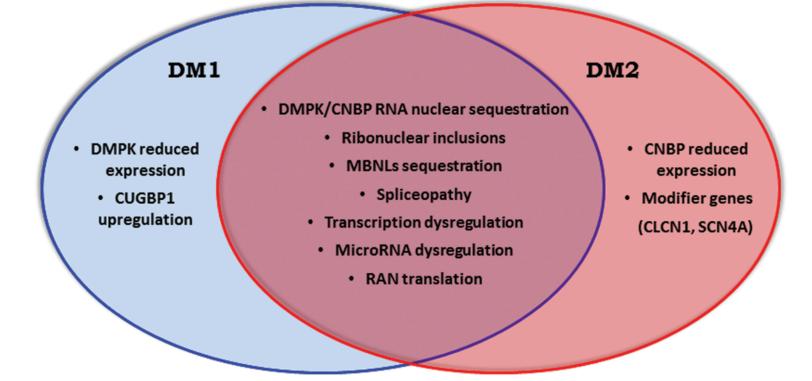


DM1 vs DM2



Journal of Neuronusculur Diseases 2 (2015) 559-571 DOI 10.3233/IND-150088 IOS Press	
Research Report	
	Meola and Cardani, 2015
Myotonic Dystrophy Type 2: An Update	
Myotonic Dystrophy Type 2: An Update on Clinical Aspects, Genetic and	
Pathomolecular Mechanism	

the phenotypic differences between DM1 and DM2 can be explained by **other cellular and molecular pathways** involved besides the shared toxic-RNA gain of function hypothesized



Alternative splicing

Differences in aberrant expression and splicing of sarcomeric proteins in the myotonic dystrophies DM1 and DM2

Arma Vihola - Linda L. Bachinski - Mario Sirito - Shodimu-Emmanuel Oluémi -Shohrae Hajilashi - Keth A. Baggerly - Olayinka Raheem - Hannu Haapasalo -Tiina Suominen - Jeanette Holmlund-Hampi - Anders Paetau - Rosanna Cardani -Giovanni Meola - Hannu Kalimo - Lars Edström - Rali Krahe - Bjarne Uid Vihola et al., 2010

DM1 vs DM2

differences in muscle gene expression and splicing: in particular, the aberrant splicing isoform of TNNT3 is twice as frequent in DM2 compared to DM1

MOREOVER

different protein expression pattern in the highly atrophic fibers has been found between DM1 and DM2

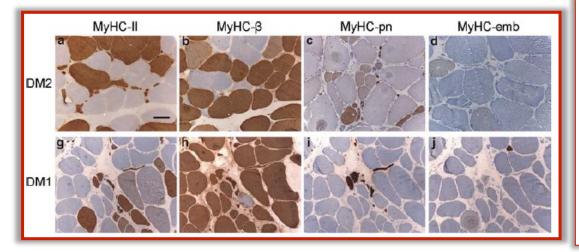


Table 3 Immunohistochemistry results of highly atrophic fibers				
Protein	Gene	DM2	DM1	
MyHC-IIa	MYH2	+++	+++	
MyHC-beta	MYH7	(+)	+++	
MyHC-pn	MYH8	+++	+++	
MyHC-emb	МҮНЗ	(+)	(+)	
fTnT	TNNT3	++	++	
NCAM	NCAMI	++	+	
Myogenin	MYOG	(+)	(+)	
Vimentin	VIM	(+)	+	

Protein expression: (+), in <1% of highly atrophic fibers; +, in 1–10%; ++, in 30–50%; +++, in >75%. The results indicate how many fibers of the highly atrophic fibers pool expressed each given antigen in DM2 (n = 20) and DM1 (n = 5) muscle biopsies

Alternative splicing

PLOS ONE



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Genome Wide Identification of Aberrant Alternative Splicing Events in Myotonic Dystrophy Type 2

Alessandra Perfetti¹⁹, Simona Greco¹⁹, Pasquale Fasanaro², Enrico Bugiardini³, Rosanna Cardani⁵, Jose M. Garcia Manteiga⁴, Michela Riba⁴, Davide Cittaro⁴, Elia Stupka⁴, Giovanni Meola^{3,3}, Fabio Martelli¹⁹ Melculu radioby Liberator, IRCS Policinko San Donato. San Donato Miance, Mian, Italy, 2 Epigenetics & Regeneative Pharmacology, IRCS Fondazione Santa

Lucia, Rome, Italy, 3 Department of Neurology, University of Milan, RCCS Policlinico San Donato, Milan, Italy, 4 Center for Translational Genomics and Bioinformatics, San Raffaele Scientific Institute, Milan, Italy, 5 Laboratory of Muscle Histopathology and Molecular Biology, IRCCS Policlinico San Donato, San Donato Milanese, Milan, Italy Perfetti et al., 2014

In DM2 muscle biopsies 273 alternative spliced exons in 218 genes were identified



many of these splicing events had been previously described as deregulated either in DM1 or DM2 or both

a subset of alternative splicing events were validated by qPCR in biceps brachii biopsies from 19 DM2 and 15 CTR age and sex matched patients

previously described in DM1 and/or DM2 PDLIM3 LIMCH1 NDUFV3 CAMK2G

ZMYND11 PDP1 ERI2 VCL MBOAT7 LAMC2

not previously described in DM1 and/or DM2

Alternative splicing



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Alessandra Perfetti¹⁹, Simona Greco¹⁹, Pasquale Fasanaro², Enrico Bugiardini², Rosanna Cardani⁵, Jose M. Garcia Manteiga⁴, Michela Riba⁴, Davide Cittaro⁴, Elia Stupka⁴, Giovanni Meola^{2,5}, Fabio Martelli¹³ ¹ Miecuk coluciolev Labatori, RICS Policinko Sin Donato, San Donato Mianeau, Mian, taly, 2 Epigenetic & Regenerative Pharmacology, RICS Fondezione Santi

Lucia, Rome, Italy, 3 Department of Neurology, University of Milan, IRCCS Policlinico San Donato, Milan, Italy, 4 Center for Translational Genomics and Bioinformatics, San Baffaele Scientific Institute, Milan, Italy, 5 Laboratory of Muscle Histopathology and Molecular Biology, IRCCS Policlinico San Donato, San Donato, Milanese, Milan, Italy Perfetti et al., 2014

the molecular pathways involving the identified aberrantly spliced genes, were studied by Interactive Pathway Analysis

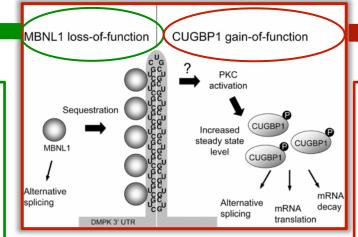
Table 1. Most significant categories and functions.	
DISEASE AND DISORDERS	p value
mmunological disease	3.116.04 2.136.02
Neurological disease	3.116.04 2.306.02
Skeletal and Muscular Oisorders	3.11E 04 1.77E 02
Cancer	9,226.04 2,466.02
Reproductive System Orsense	9.22E 04 1.77E 02
MOLECULAR AND CELLULAR FUNCTIONS	p value
Cell Oeath and Surwyal	1.05E 04 2.13E 02
Cellular Oevelopment	1.81E 04 1.83E 02
Cell Marphalogy	2.83E 04 1.77E 02
Cellular Movement	3.93E 04 2.48E 02
Cell To Cell Signaling and Interaction	1.25E 03 2.46E 02
PATHWAYS	logip value)
Lanosterol Biosyntheas	1.75600
Netrin Signaling	1.73600
Epithelial Adherens Junction Signaling	1.62500
Fatty And Biosynthesis Initiation II	1.46600
Palmitate Brosynthesis I (Animals)	1.46600
Jrea Cycle	1.48600
Caloum Signaling	1.42600
TO P CARDIOTOXIC FUNCTIONS	p value
noreased Levels of Albumin	1.77E 02 1.77E 02
noreased Levels of Alkaline Phosphatase	1.77€02 6.12€01
Cardiac Anythma	4.07E03 3.25E01
fachycardia	4.07E 03 3.25E 01
Cardiac Oilation	1.77E 02 1.43E 01
Congenital Heart Anonialy	1.77E 02 433E 01
Cardiac Hypoplasa	3.93€ 02 3.93€ 02

The affected genes are involved in numerous pathways and networks important for muscle physio-pathology, suggesting that the identified variants may contribute to DM2 pathogenesis.



of particular interest Skeletal and Muscular Disorders Neurological Diseases Cell Death and Survival Cellular Development Calcium signaling Cardiac Arrythmia

CUGBP1 expression

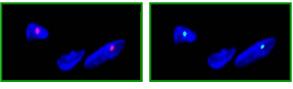


CUGBP1 overexpression has been clearly demonstrated in DM1 but not in DM2 muscle

conflicting data have been reported on the expression of CUGBP1 in DM2 human skeletal muscle

both in DM1 and DM2

it is clear that MBNL1 is depleted from nucleoplasm through recruitment into ribonuclear inclusions even when clinical symptoms and muscle alterations are very mild

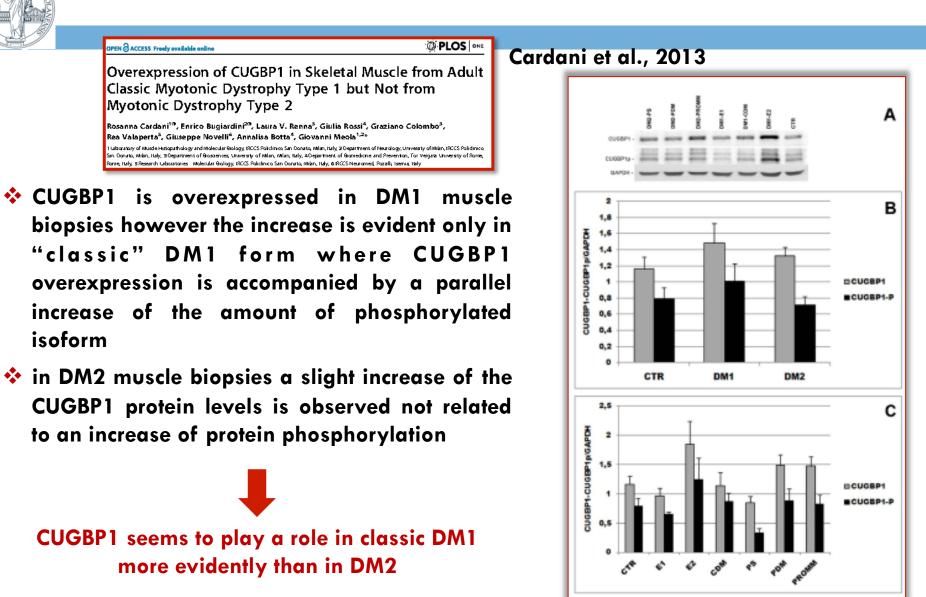


Toxic RNA

MBNL1 foci

CUGBP1





ZNF9/CNBP



PLOS ONE PEN access Freely available online Cardani et al., 2013 Overexpression of CUGBP1 in Skeletal Muscle from Adult Classic Myotonic Dystrophy Type 1 but Not from Myotonic Dystrophy Type 2 DM2-PROM M2-PROM DM2-PDM DM1-E2 DM1-E2 DM2-PS Rosanna Cardani¹⁰, Enrico Bugiardini²⁰, Laura V. Renna³, Giulia Rossi⁴, Graziano Colombo³, А CTR Rea Valaperta⁵, Giuseppe Novelli⁶, Annalisa Botta⁴, Giovanni Meola^{1,2}* 1 laboratory of Musde Histopathology and Molecular Biology, IRCCS Policinico San Conarto, Milan, Italy, 2 Oepartment of Neurology, University of Milan, IRCCS Policinico Sin Donato, Milan, Haly, 3 Department of Bosoences, University of Milan, Milan, Haly, 4 Department of Bonnedicine and Prevention, Yor Vergata University of Some, Rome, Italy, 5 Research Laboratories - Molecular Biology, IRCCS Policinico San Conato, Milan, Italy, 6 RCCS Neuromed, Pozalili, Isemia, Italy ZNF9/CNBP GAPDH в 1,2 ** 1 ZNF9/CNBP protein levels are significantly ZNF9/GAPDH 0, 0, 0, 10, 200 reduced in DM2 muscle biopsies compared to DM1 and non-diseased biopsies 0,2 0 CTR DM1 DM2 1,2 С ZNF9/CNBP expression might play a role in phenotypic differences between DM1 and ZNF9/GAPDH 0,8 **DM2** 0.6 0.4 0,2 o

CTR

E1

E2

CDM

PS

PDM PROMM

MicroRNA



OPEN © ACCESS Freely available online PLOS one Deregulated MicroRNAs in Myotonic Dystrophy Type 2 Giovanni Meola^{1,2}, Fabio Martelli^{1,8} Simona Greco¹⁰, Alessandra Perfetti¹⁰, Pasquale Fasanaro³, Rosan na Cardani¹, Maurizio C. Capogrossi³, Giovanni Meola^{1,2}, Fabio Martelli^{1,8} 1/8CS Relidinco Sin Donato, Milan, M

MiRNA profiling identified 20 miRNAs significantly modulated in DM2 muscle compared to CTR

PROFILING				
CTR DM2		CTR DM2 ^{miR}		р
		miR-381	5.1	*
		miR-34a-5p	3.9	*
		miR-34c-5p	3.6	**
		miR-146b-5p	3.6	**
		miR-133b	3.3	
		miR-323a-3p	3.0	*
		miR-208a	2.9	**
		miR-221-3p	2.3	*
		miR-34b-3p	2.1	*
		miR-376a-3p	2.1	*
		miR-432-5p	2.0	*
		miR-410	2.0	*
		miR-517a-3p	-3.3	*
		miR-383	-2.9	٠
		miR-520h	-2.8	**
		miR-193b-3p	-2.5	••
		miR-133a	-2.3	**
		miR-378a-3p	-2.2	**
		miR-518e-3p	-2.1	*
		miR-125b-5p	-2.0	*
-3.0 +3.0				

validation by more sensitive and specific qPCR assays identified 11 deregulated miRNAs

miRNA score allowed to discriminate DM2 patients from CTR with a good sensitivity and specificity.

		DM2		DM1	
DM2 DM1	miR	FOLD CHANGE	р	FOLD CHANGE	р
	miR-221-3p	6.5	***	1.2	ns
	miR-34c-5p	5.7	***	2.0	ns
	miR-208a	4.6	***	2.4	
	miR-381	4.3	***	2.8	•
	miR-34b-3p	4.0	***	1.8	ns
	miR-34a-5p	3.2	***	1.3	ns
	miR-146b-5p	2.1	•	1.3	ns
	miR-193b-3p	-6.1	•	-1.6	**
	miR-125b-5p	-5.7	***	-1.4	ns
	miR-378a-3p	-3.7	**	-1.0	ns
	miR-193a-3p	-2.6		1.3	ns

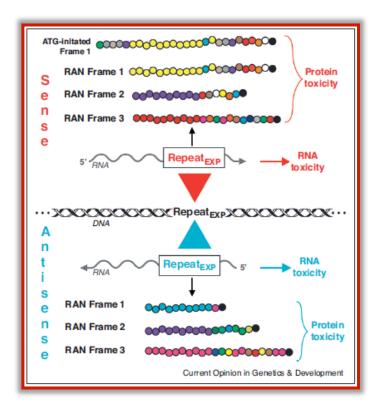
miR-193b-3p, miR-208a and miR-381 showed a similar significant modulation also in DM1 patients

RAN Translation

Non-ATG-initiated translation directed by microsatellite expansions

Tao Zu^{a,b,c}, Brian Gibbens^{a,b,c,1}, Noelle S. Doty^{a,b,c,1}, Mário Gomes-Pereira^d, Aline Huguet^d, Matthew D. Stone^{e,f}, Jamie Margolis^{a,b,c}, Mark Peterson⁹, Todd W. Markowski^{e,f}, Melissa A. C. Ingram^{a,b,c}, Zhenhong Nan^h, Colleen Forsterⁱ, Walter C. Low^h, Benedikt Schoserⁱ, Nikunj V. Somia^{a,b}, H. Brent Clark^{c,i,k}, Stephen Schmechelⁱ, Peter B. Bitterman⁹, Geneviève Gourdon^d, Maurice S. Swanson¹, Melinda Moseley^{a,b,c}, and Laura P. W. Ranum^{a,b,c,2,3}

Zu et al., 2010



a repeat expansion mutation can produce potentially toxic RNA and protein products expressed through a combination of:

- bidirectional transcription
- ATG-initiated translation
- repeat associated non-ATG (RAN) translation

RAN translation of the expanded repeat results in the expression of up to six distinct RAN proteins

In vitro and	In vitro and in vivo evidence for RAN translation					
	Repeat	In Vitro evidence of RAN proteins	In Vivo evidence of RAN proteins	Reference		
SCA8	CAG+CTG	Gin _S ^{a,b,g} , Ala _S ^{a,b,c,d,f,g} , Ser _S ^{a,b,g} Leu _{AS} ^a , Ala _{AS} ^a , Cys _{AS} ^a	Ala _s ^{j, m}	Zu et al. [12**]		
DM1	CAG•CTG	GIn _{AS} ^{a,e,f} , Ala _{AS} ^a , Ser _{AS} ^a	GIn _{AS} ^{i,j,l,m}	Zu et al. [12**]		
FXTAS	CGG+CCG	Glys ^a , Alas ^{a,d,g}	Glys ^{h,j,m}	Todd et al. [24**]		
C9ORF72 ALS FTD	$G_4C_2 \bullet G_2C_4$	GlyPro _S ^e , GlyAla _S ^e	GlyPro _{S/AS} ¹ , GlyAla _S ¹ , GlyArg _S ¹ GlyPro _{S/AS} ¹ GlyPro _{S/AS} ^k GlyAla _S ¹	Mori et al. [47**] Ash et al. [44**] Almeida et al. [36] Mackenzie et al. [45		
		GlyPro _{S/AS} ^e , ProArg _{AS} ^e	GlyPro _{S/AS} ^I , ProArg _{AS} ^I , ProAla _{AS} ^I GlyPro _{S/AS} ^k GlyArg _S ^I , GlyAla ^m ProArg _{AS} ^I , ProAla _{AS} ^I	Gendron et al. [38*] Donnelly et al. [37*] Mori et al. [46*]		
		GlyPro _{S^{a,f}, GlyArgs^{a,e,f} GlyAla^{a,f} GlyPro_{S/AS}^{a,e,} ProArg_{AS}^{a,e,f}, ProAla_{AS}^{a,e,f}}	GlyPros ^{l.m} , GlyArgs ^{l.m} , GlyAla ^m GlyPro _{AS} ^l , ProArg _{AS} ^{l.m} , ProAla _{AS} ^{l.m} GlyArg _S ^l , GlyAla _S ^l GlyPro _{S/AS} ^l , ProArg _{AS} ^l , ProAla _{AS} ^l	Zu et al. [48**] Mann et al. [49]		

RAN Translation



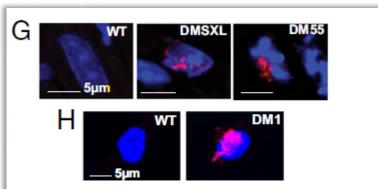
Non-ATG-initiated translation directed by microsatellite expansions

Tao Zu^{a,b,c}, Brian Gibbens^{a,b,c,1}, Noelle S. Doty^{a,b,c,1}, Mário Gomes-Pereira⁴, Alime Huguet⁴, Matthew D. Stone^{s,f}, Jamie Margolis^{a,b,c}, Mark Peterson⁹, Todd W. Markowski^{e,f}, Melissa A. C. Ingram^{a,b,c}, Zhenhong Nan¹, Colleen Forsterⁱ, Walter C. Low^h, Benedikt Schoser^j, Nikunj V. Somia^{a,b}, H. Brent Clark^{c,i,k}, Stephen Schmechel^j, Peter B. Bitterman⁹, Geneviève Gourdon⁴, Maurice S. Swanson¹, Melinda Moseley^{a,b,c}, and Laura P. W. Ranum^{a,b,c,2,3}

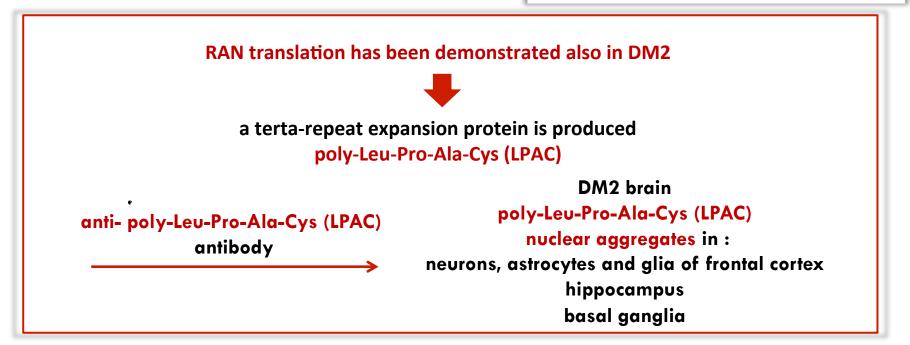
In DM1 mouse model polyGln nuclear aggregates

- Cardiac myocytes
- leukocytes
- myoblasts
- skeletal muscle

anti- poliGln antibody



Zu et al., 2010



Myoblast senescence

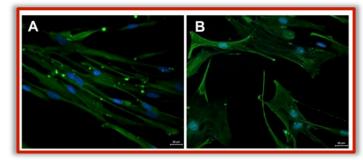
European Journal of Histochemistry

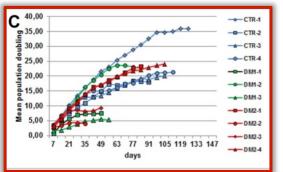
Premature senescence in primary muscle cultures of myotonic dystrophy type 2 is not associated with p16 induction

L.V. Renna,¹ R. Cardani,² A. Botta,³ G. Rossi,³ B. Fossati,⁴ E. Costa,^{5,6} G. Meola^{2,4}

Renna et al., 2014

DM myoblasts have lower proliferative capability than control myoblasts and reach *in vitro* senescence earlier than controls

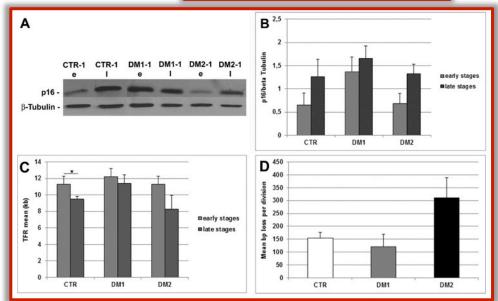




HOWEVER

differentely from DM1, the p16 pathway is not responsible for the premature growth arrest observed in DM2 myoblasts which stop dividing with telomeres shorter than controls

These data could explain the different histological alterations observed between DM1 and DM2 skeletal muscle as for example the selective type 2 fiber atrophy present in DM2 muscle





Modifier genes

Myotonia

In DM2 patients:

- usually is less severe than in DM1 patients
- sometimes may be difficult to reveal even with EMG

however

in several DM2 patients it can be very severe

in a cohort of 45 genetically confirmed DM2 patients 4/45 patients (8,89%) presented a severe or early onset myotonia.

The genetic analysis of CLCN1 and SCN4A revealed that

- 2 patients showed a recessive mutation in CLCN1 gene
- 2 patients showed a mutation in SCN4A gene

Modifier gene: CLCN1



J Neurol DOI 10.1007/s00415 012 6462 1

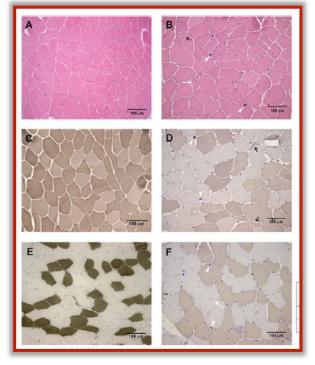
ORIGINAL COMMUNICATION

Co-segregation of DM2 with a recessive CLCN1 mutation in juvenile onset of myotonic dystrophy type 2

Rosanna Cardani · Marzia Giagnacovo · Annalisa Botta · Fabrizio Rinaldi · Alessandra Morgante · Bjarne Udd · Olayinka Raheem · Sini Penttilä · Tiina Suominen · Laura V. Renna · Valeria Sansone · Enrico Bugiardini · Giuseppe Novelli · Giovanni Meola

Cardani et al., 2012

A 15-year-old DM2 patient and her mother were studied to further investigate the unusually young onset in this DM2 family



 the age at onset was earlier in the daughter than in the mother
the daughter's clinical, histopathological and biomolecular findings did not show greater severity than those observed in her mother

HOWEVER

daughter presented handgrip myotonia at the age of 14 years.

Direct sequencing CLCN1 gene revealed a heterozygous mutation c.501C>G p.F167L in daughter maternally inherited

the co-segregation of DM2 with a recessive CLCN1 mutation provided the explanation for the unusual clinical findings

Modifier gene: SCN4A

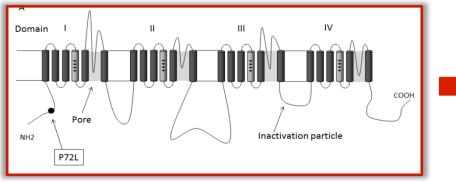
SCN4A mutation as modifying factor of Myotonic Dystrophy Type 2 phenotype

E. Bugiardini ^{a,1}, I. Rivolta ^{b,1}, A. Binda ^b, A. Soriano Caminero ^c, F. Cirillo ^d, A. Cinti ^e, R. Giovannoni ^e, A. Botta ^f, R. Cardani ^g, M.P. Wicklund ^c, G. Meola ^{a,g,*} Bugiardini et al., 2015

A 26 year old patient complaining of hand cramps and difficulty relaxing her hands after activity was evaluated

Genetic testing was positive:

- for DM2 (2650 CCTG repeat)
- for a variant c.215C>T (p.Pro72Leu) in the SCN4A gene



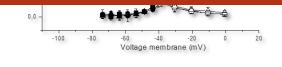
The variation affects the cytoplasmic Nterminus domain of Nav1.4, where mutations have never been reported



Electrophysiological studies of the P72L variant

If CLCN1 screening is negative, this case supports for screening SCN4A mutations in DM2 patients with atypical cases with severe myotonia

increase cell excitability





Myotonic dystrophies management

DM1

DM2

Brain	Psychological, educational, and counseling evaluations as needed Structural imaging as required Routinely assess for sleep disturbances and respiratory insufficiency	Psychological, educational, and other counseling treatment and services CNS medications (for example, stimulants) as necessary under close supervision of care providers
Heart	Yearly electrocardiograms Cardiology consultation for symptomatic patients and long-term follow-up care	Prompt pacemaker placement as needed
Respiratory	Serial monitoring of sitting and supine respiratory function; including forced vital capacity Polysomnography and pulmonary medicine consultation as required	Yearly immunizations Noninvasive or invasive ventilation as required Serial evaluation by pulmonary medicine and sleep consultation as required
Anesthesia	Before elective surgery, have anesthesia consultation and pulmonary medicine evaluation ECG reviewed by cardiology consult Discuss known risks and any previous anesthesia related problems	Use of regional anesthesia over general when appropriate Use of non-depolarizing muscle relaxants Reduce use of opioids In general anesthesia, protection of the airway and minimizing aspiration, careful cardiac monitoring, and extensive postoperative monitoring (at least 24 hours)



Myotonic dystrophies management

DM1

DM2

Hypersomnia and fatigue	Polysomnograms Metabolic and endocrine screens Psychological, educational, and sleep consultant evaluations	Use of continuous positive airway pressure (CPAP) or bi- level positive airway pressure (BiPAP) Use of CNS stimulants
Endocrine	Symptomatic assessment of testosterone deficiency Yearly lipid profile, thyroid screening, diabetes screening Monitor sleep disturbances	Hormone replacement as required Dietary intervention Medications for lipid and glucose control as needed Treatment for sleep disturbances as required
Gastrointestinal	Occupational and physical therapy consultation (dysphagia) Metabolic and endocrine screens Dietician, gastrointestinal consultations Careful assessment of bloating and signs of pseudo-obstruction	Gastroesophageal reflux may be treated with avoidance of late-night meals, elevation of the head of the bed, and medications Constipation, diarrhea, abdominal pain, and bloating may be treated with modifying the diet to small, low-fat meals Surgery as appropriate for gall bladder disease Use of cholestyramine may help alleviate diarrhea
Pregnancy	Obtain obstetrician and genetic consultation prior to pregnancy as appropriate Discuss possible complications Coordinate monitoring of pregnancy with other care providers, including a neonatal pediatric specialist Closely monitor respiratory function during the third trimester	During delivery, monitor mother's ECG Use regional anesthesia Notify consultants of mother's status and request urgent evaluations as necessary



RNA level: small molecules

Selective inhibition of MBNL1–CCUG interaction by small molecules toward potential therapeutic agents for myotonic dystrophy type 2 (DM2)[†]

Chun-Ho Wong, Yuan Fu, Sreenivasa Rao Ramisetty, Anne M. Baranger* and Steven C. Zimmerman* 2011

Small Molecules that Target the Toxic RNA in Myotonic Dystrophy Type 2

Lien Nguyen, JuYeon Lee, Chun-Ho Wong, and Steven C. Zimmerman*[a]

2014

Structure of the Myotonic Dystrophy Type 2 RNA and Designed Small Molecules That Reduce Toxicity

2014

Jessica L. Childs-Disney^{#a}, Ilyas Yildirim^{#b}, HaJeung Park^{a,c}, Jeremy R. Lohman^a, Lirui Guan^a, Tuan Tran^a, Partha Sarkar^d, George C. Schatz^b, and Matthew D. Disney^{a,*}

Most of the molecules identified resulted to be toxic in cellular assays



Take home message



Take home message

The enormous advances in the understanding of the molecular pathogenesis of DM1 and DM2 has revealed pathways of molecular pathogenesis more complex than previously appreciated

however

the basis for the differences between DM1 and DM2 has not been clarified at the molecular level

important for the development of effective therapies

Thanks to.....

Medical doctors



Biologists

Prof. Giovanni Meola Dr. Giuseppe Rotondo Dr. Barbara Fossati Dr. Enrico Bugiardini Dr. Mauro Toffetti Dr. Elisa Brigonzi Dr. Michele Cavalli Dr. Lorenzo Saraceno

Dott.ssa Rosanna Cardani

Dott.ssa Laura V Renna Dott.ssa Rea Valaperta Dott.ssa Francesca Bosè



.....and to patients and their families



Università degli Studi di Milano

